# A Systematic Approach to Developing Stability Indicating Methods (SIMs)



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# Outline

- Purpose
- Regulatory Requirements
- Development Steps
- Method Acceptability
- Case Study



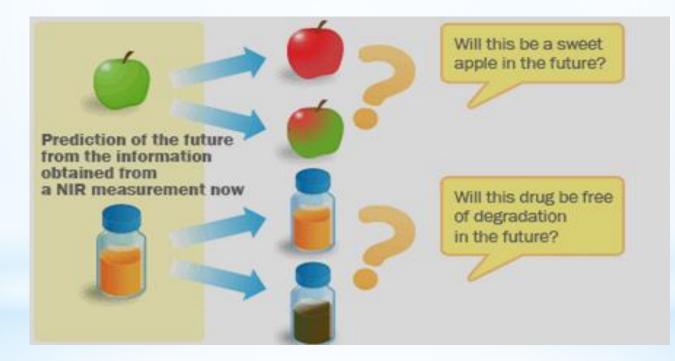
## **Purpose of SIM**

- 1970s, rising concerns over drug product stability
- 1975, USP added drug product expiration dating
- 1987, FDA issued guidelines on submission of stability data
- 2009, WHO issued its own guidelines



### **Purpose of SIM**

### Necessary to establish drug substance or drug product stability over shelf-life





## **Regulatory Requirements**

- -multiple guidance documents (see references)
- -new definition as of July 2015
- -some detail provided
- -what constitutes SIM open
  - for interpretation





# Regulatory Definition (August 2000)

"accurately measures the active ingredients, without interference from degradation products, process impurities, excipients, or other potential impurities."

Guidance for Industry, Analytical Procedures and Method Validation, U.S. Department of <u>Health</u> and Human Services FDA, August 2000



# **New Definition (July 2015)**

"If a procedure ...can detect changes in a quality attribute(s) of the drug substance and drug product during storage, it is considered a stability-indicating test."

Guidance for Industry, Analytical Procedures and Methods Validation for Drugs and Biologics, U.S. Department of Health and Human Services FDA, July 2015



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## **New Definition (July 2015)**

- Specificity more detail given
  - Samples spiked with target analytes and all known interferences
  - Samples that have undergone various laboratory stress conditions
  - Actual product samples (produced by the final manufacturing process) that are aged or stored under accelerated temperature/ humidity conditions



### **Development Steps**

- Analytical Methodology
- Method Development Strategy
- Current Best Practices





### **Development Steps**

### Analytical Methodology





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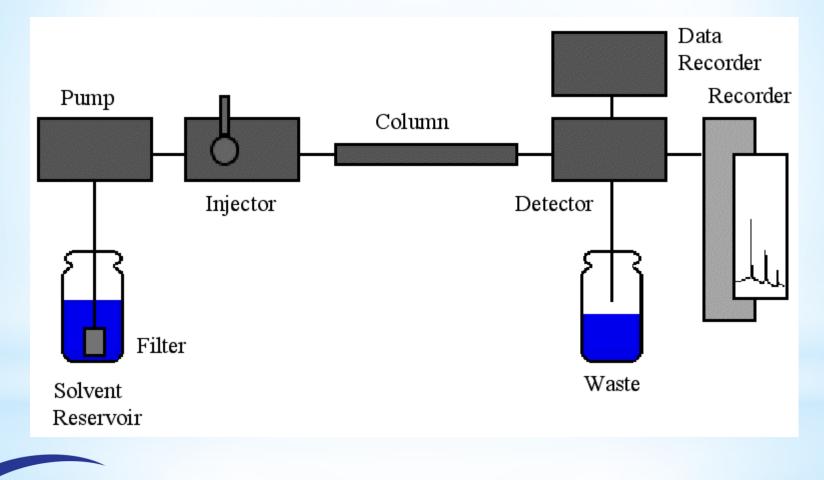
**Examples** 

HPLC/UPLC LCMS GCMS **TLC/HPTLC** CE **UV** spectroscopy



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### **Majority of SIMS are HPLC**



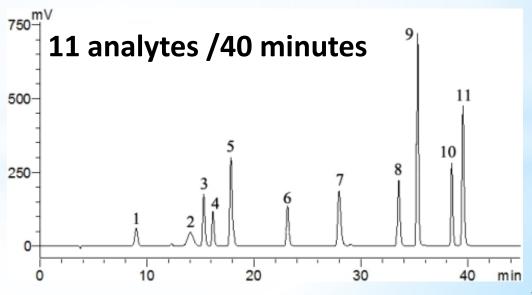


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# **HPLC Advantages**

- For impurity profiling power of separation
- Good sensitivity / multiple detectors
- Diverse range of analytes
- Continuous innovations
- Automated

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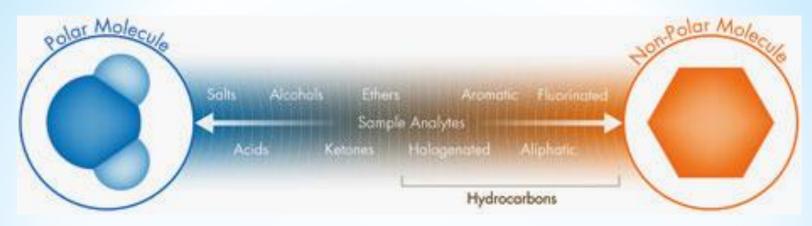
### **HPLC Advantages**

Multiple modes of separation base on chemical characteristics of compound: **Polarity Electrical Charge Molecular Size** 





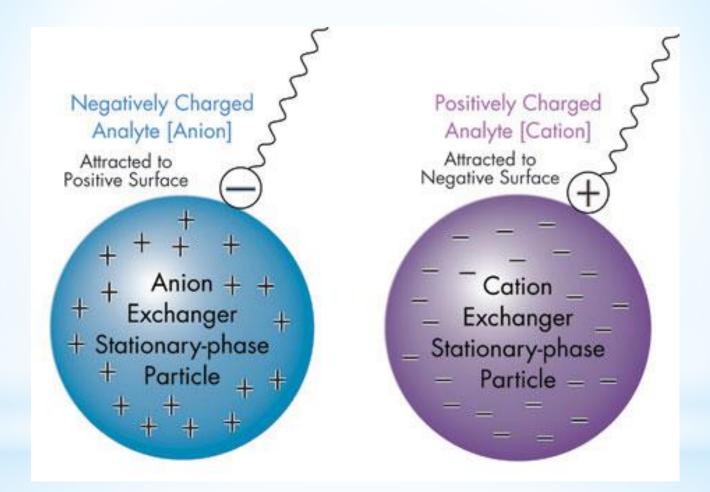
# **Polarity**



## Reverse Phase Normal Phase Hydrophobic Interaction (HIC) Hydrophilic Interaction (HILIC)

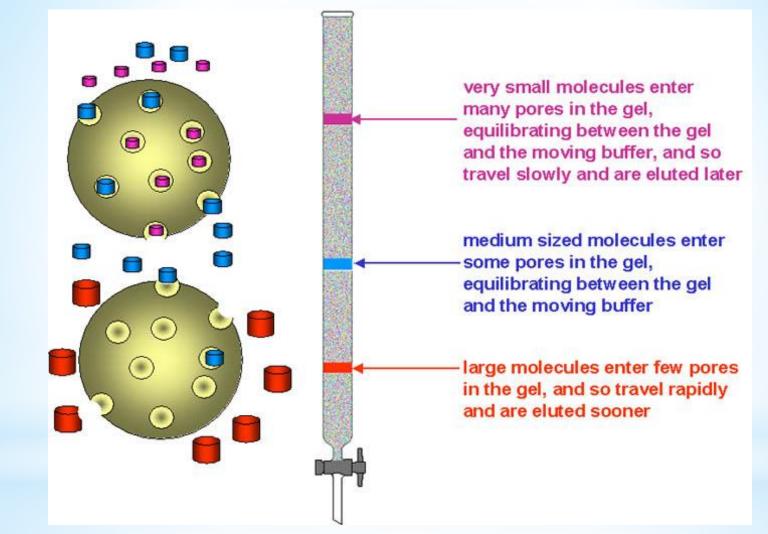


### Ion-Exchange





### **Size Exclusion**

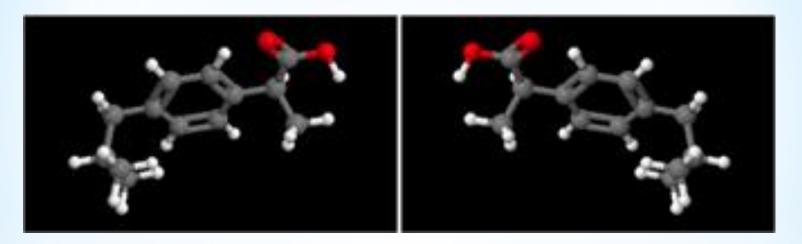


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# **Chiral chromatography**

#### -well known example is ibuprofen



S+ibuprofen (left) and R-ibuprofen (right) showing their mirror image relationship



## **Chiral chromatography**

**Chiral stationary phase. Requires 3** interactions through: **H-bonding**  $\pi$ - $\pi$  interactions **Dipole stacking Inclusion complexing Steric bulk Conducted** in both normal / reverse phase modes



# 75% of all HPLC methods are Reverse Phase



## **Reverse Phase HPLC**

# Definition: mobile phase is more polar than stationary phase. Stationary phase hydrophobic.



## **Selectivity in Reverse Phase HPLC**

## Three equilibria involved:

- 1. Solute interaction with mobile phase
- 2. Solute interaction with solid phase
- 3. Solute interaction with support

### **Selectivity in Reverse Phase HPLC**

**Molecular interactions** 

- -Dispersion /hydrophobic
- -Charge Transfer  $(\pi \pi)$  / aromatic
- -Hydrogen bonding / acidic
- -Dipole-dipole / basic



### **Reverse Phase HPLC**

# Detection Method Stationary Phase Isocratic or Gradient Mode



### **RP HPLC Detection**

- Most popular UV/Vis, diode array
- Mass Spectrometry increasing use
- Additional refractive index (RI), fluorescence, charged aerosol detector (CAD), evaporative light scattering (ELSD), amperometry, and conductivity



## **UV/Vis Detection**

#### **Example Chromophores**

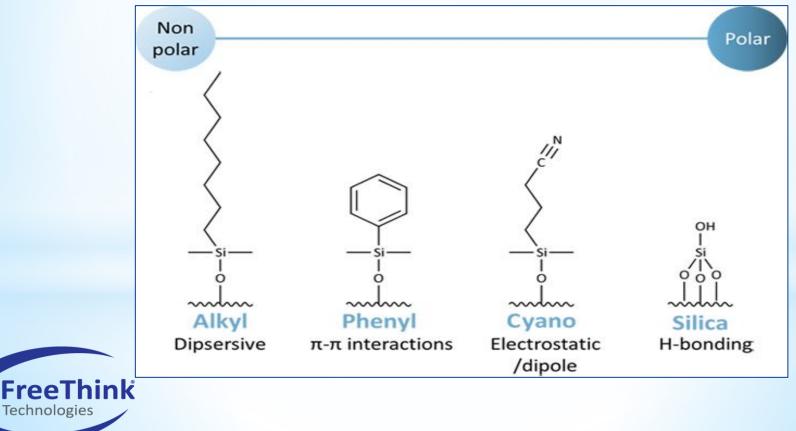
| C=C<br>C=C<br>C=O | Example       | Transition  | $\lambda_{max}/nm$ | €/mol <sup>−1</sup> m <sup>2</sup> |
|-------------------|---------------|---|--------------------|------------------------------------|
|                   | ethylene      | $\pi \rightarrow \pi^*$   | 165                | 1500                               |
|                   | acetone       | $\begin{array}{c} \pi \rightarrow \pi^* \\ n \rightarrow \pi^* \end{array}$ | 188<br>279         | 90<br>1.5                          |
| -N=N-             | azomethane    | $n \rightarrow \pi^*$   | 347                | 0.45                               |
| -N=0              | nitrosobutane | $\pi \to \pi^{*}$   | 300                | 10                                 |
|                   |               | $n \rightarrow \pi^*$   | 665                | 2                                  |
|                   | benzene       | $\pi \to \pi^*$   | 200<br>255         | 800<br>21.5                        |

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# **Stationary Phase**

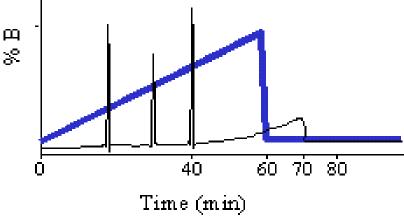
- Variety phases: alkyl, fluoroalkyl, amide, amine, cyano, diol, phenyl, etc.
- Most widely used is C18 alkyl phase



## **Isocratic or Gradient Elution**

**Isocratic**: composition of mobile phase remains constant throughout the run. Most used for assay, dissolution. Separating 2 or 3 compounds in a single run.

**Gradient**: Mobile phase composition varies over run. Most used for impurity profiling and degradation studies.





### **Development Steps**

### Method Development Strategy





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### **Method Stages**

- Pure API
- Changing API process
- Changing salt form
- Formulation concerns
- It is transferable?



# **Method Development**

### **Starting considerations:**

- Assay only, or assay & related substances? Multiple actives?
- Special sample type such as inorganic ions, enantiomers, biomolecules, polymers, carbohydrates, isomers?
- Is it feasible to determine all related substances in one method?



### **Method Development**

- 1. Gather information on chemical & physical properties of analytes
  - Literature search
  - Synthetic process
  - pKa (pH 50% of compound protonated) and log P (lipophilicity) values
  - Check availability of method
    development software (DryLab)



#### **2.** Evaluate analyte mixture

- Range of pKa and log P values
- Determine critical pairs
- Presence of amine-containing compounds (tailing)
- Presence of amphoteric compounds (presence of both acidic and basic functional groups)



#### **3. Develop starting conditions**

- Based on step 2, select 3-4 stationary phases (commercial kits are available)
- Start at low pH
- Run gradient 5 -100% over 20-40min
- Use diode array detection for λmax values
- Use test mixtures of critical pairs



### Selectivity

Changed when we change the chemistry of chromatographic system

Variables that affect selectivity:

-solvent strength and type

- -temperature of the column
- -gradient steepness
- -buffer and other additives

-type of column packing



### **Organic Solvents**

| Solvent<br>Water<br>Methanol<br>Acetonitrile<br>THF<br>IPA<br>Hexane | UV cut off (nm)<br>200<br>205<br>190<br>215<br>210 | Viscosity (mPa s)<br>1.00<br>0.6<br>0.37<br>0.55<br>2.35 | B.P. (°C)<br>100<br>65<br>82<br>65<br>83 | Polarity (P') <sup>1</sup><br>9.0<br>6.6<br>6.2<br>4.2<br>4.3 |     |       |    |   |
|--|--|--|--|---|-----|-------|----|---|
|  |  |  |  |   | 195 | 0.313 | 69 | 0 |



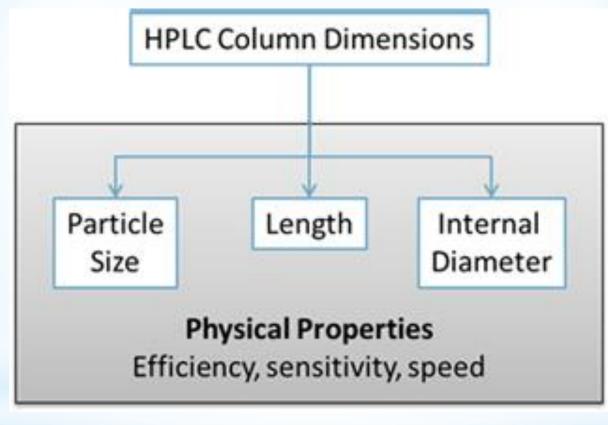
#### **Common Buffers**

| Buffer                       | р <i>К</i> _ | pH range    | UV cut off (nm) |
|------------------------------|--------------|-------------|-----------------|
| Phosphate, pK 1              | 2.1          | 1.1 - 3.1   | <200            |
| Phosphate, pK 2              | 7.2          | 6.2 - 8.2   | <200            |
| Phosphate, pK <sub>a</sub> 3 | 12.3         | 11.3 - 13.3 | <200            |
| Citrate, pK 1                | 3.1          | 2.1 - 4.1   | 230             |
| Citrate, pK 2                | 4.7          | 3.7 - 5.7   | 230             |
| Citrate, pK <sub>a</sub> 3   | 6.4          | 5.4 - 7.4   | 230             |
| Carbonate, , pK 1            | 6.1          | 5.1 - 7.1   | <200            |
| Carbonate, pK <sub>a</sub> 2 | 10.3         | 9.3 - 11.3  | <200            |
| Formate                      | 3.8          | 2.8 - 4.8   | 210 (10 mM)     |
| Acetate                      | 48           | 3.8 - 5.8   | 210 (10 mM)     |
| Ammonia                      | 9.3          | 8.3 - 10.3  | 200 (10 mM)     |
| Borate                       | 9.2          | 8.2 - 10.2  | N/A             |
| TFA                          | 0.5          |             | 210 (0.1%)      |

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### **Column Dimensions**



#### HETP = L / N



### **Column Dimensions**

| Column Length, L (mm) | Particle Size, d <sub>P</sub> (µm) | Resolution Capacity L/d <sub>p</sub> |
|-----------------------|------------------------------------|--------------------------------------|
| 300                   | 10                                 | 30,000                               |
| 150                   | 5                                  | 30,000                               |
| 100                   | 3                                  | 33,300                               |
| 50                    | 1.7                                | 29,500                               |
| 100                   | 1.7                                | 58,820                               |
| 150                   | 1.7                                | 88,230                               |



#### 4. Optimization

- Most time-consuming step
- Speed vs. resolution
- Early stage development an iterative process that is repeated several times to accommodate unexpected impurities that arise



### **5. Method merits**

- Resolution between peaks of 1.0 or better (API peaks usually broad due to their higher concentration in sample)
- Peak shape (tailing factor 0.9-1.5) & plate number (>4000)
- Repeatability RTs ±0.1 min and R<sub>s</sub> ±0.2 units



### **5. Method merits**

- No interference from blank
- No interference from sample matrix
- Linear response for all degradants
- Able to achieve desired limit of quantitation (LOQ) for degradants
- Able to recover API & degradants from sample matrix



# **Sample Extraction Method**



# **Development Steps**

### Current Best Practices





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# **Current Best Practices**

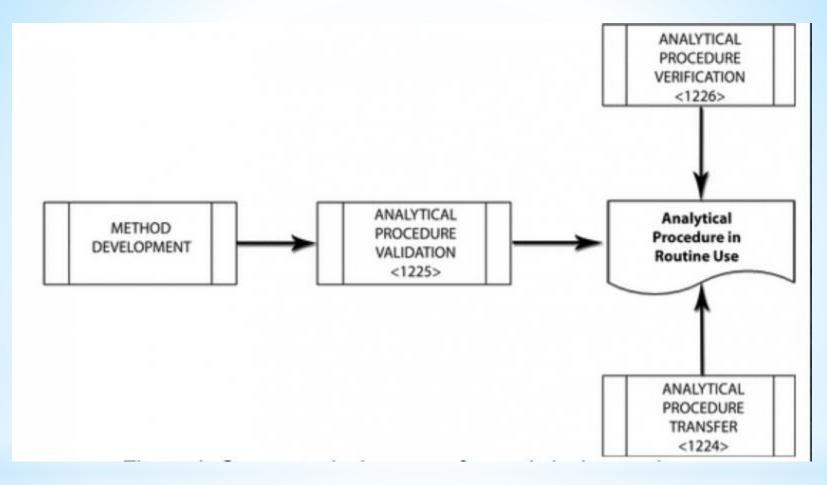
- Thorough understanding of drug substance chemistry - vulnerability assessment
- Degradation target range of 5-10%; careful not to under or over-degrade sample
- Perform "complete" identification of unknowns that exceed ICH thresholds



## **Current Best Practices**

- Pre-qualify method to ensure it is "validatable"/ method development is not GMP- want successful GMP testing
- Phase-specific strategy for validation
- Adopt USP "life cycle management" approach to SIM





#### **Typical Process**





#### Lifecycle Approach



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#### Goal

Develop single method for assay and related substances for monolayer tablet

- Assay 2 actives in tablet
- 9 related substances



### History

- Prior method developed not working after transfer
- Reported problems reported were column failures, excessive back pressure, shifting retention times, loss of resolution, difficulty obtaining LOQ



### **Simple method corrections**

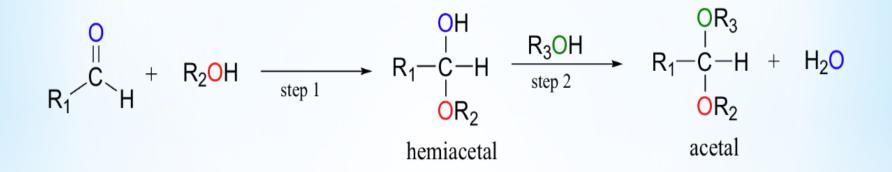
- Appropriate wavelength for detection
- Type of detector and path length
- Buffer mobile phase (stay within buffering range)
- Select column suitability for high aqueous starting conditions



- -Dual therapy product: separating 2 actives and all associated related substances (14 compounds total)
- -Not all compounds neutral, ionizable groups (pKa values should be evaluated
- -Several amine-containing compounds (peak tailing is a concern)



-Ketal chemistry a concern for 3 of the compounds (better to use aprotic solvent)





#### **Evaluation**

- pKa values: 3.98, 8.2, 9.6, 10.6
  -available pH 2.5, 5.5-7, high pH
- Log P: wide range from -0.2 to 4
- All compounds contain phenyl ring



#### **Evaluation**

- Two critical pairs with similar structures
- Target specification levels of 0.3%
- Large difference in levels of actives;
  8 mg active 1 / 90 mg active 2
- One degradant has isomer



### **Rules of thumb retention behavior:**

- Increases as #carbon atoms increases
- Branched-chain compounds elute more rapidly
- Decreases with increasing unsaturation
- Order of elution generally follows:

aliphatics > weak Lewis bases (esters, aldehydes, ketones) > strong Lewis bases (amines) > weak Lewis acids (alcohols, phenols) > strong Lewis acids (carboxylic acids)



Establish starting separation conditions: HPLC column needs to:

- operate under highly aqueous (avoid hydrophobic collapse)
- work for mixture neutral, acidic, basic compounds
- prevent tailing (do not want to add ion-pairing reagent, if possible)



Column dimensions – 4.6 mm x 25 cm, 5µm Both columns designed for high aqueous

- Supelcosil ABZ plus alkyl amide phase for mixtures of neutral, acidic, basic compounds, use with low ionic strength buffers without need for ion-suppressing modifier
- ACE C18AR C18-phenyl phase extra resolving power with π-π selectivity of the phenyl functionality, highly inert for amines



#### **Results of screening exercise**

- -50% of compound eluted in first 20 minutes
- -50% of compounds eluted after 40 minutes
- -20 minutes of "dead time"



# Results of screening exercise -at low pH, all retention times decreased, one compound elutes in void volume of column



#### **Results of screening exercise**

## -pH > 6.0, lack of resolution between critical pair below that differs only in the addition of a hydroxyl group



### **Final separation conditions:**

- Buffer: 20 mM ammonium acetate @ pH 5.8 vs. prior 100 mM sodium phosphate @ pH 6.1 (actually buffering)
- Based on UV/Vis spectra collected using PDA – wavelength set at 229 nm vs. 280 nm which was *not* detecting one of related substances



### **Final separation conditions:**

- 3. Organic modifier acetonitrile for best peak shape
- 4. Incorporated 2 gradients into method using of a step function:

-100% MP A – 75% MP A

-step to 60% MP A

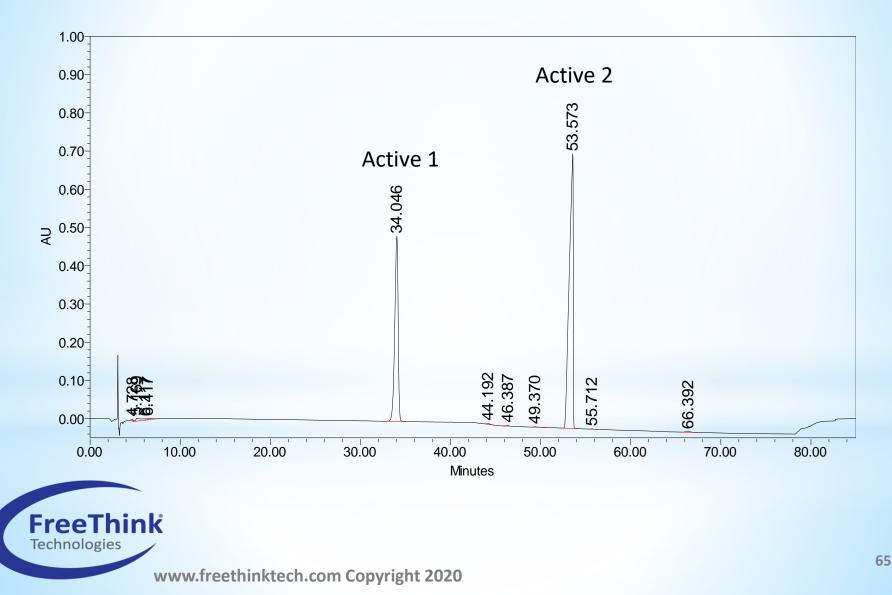
<u>-60% MP A – 0% MP A</u>

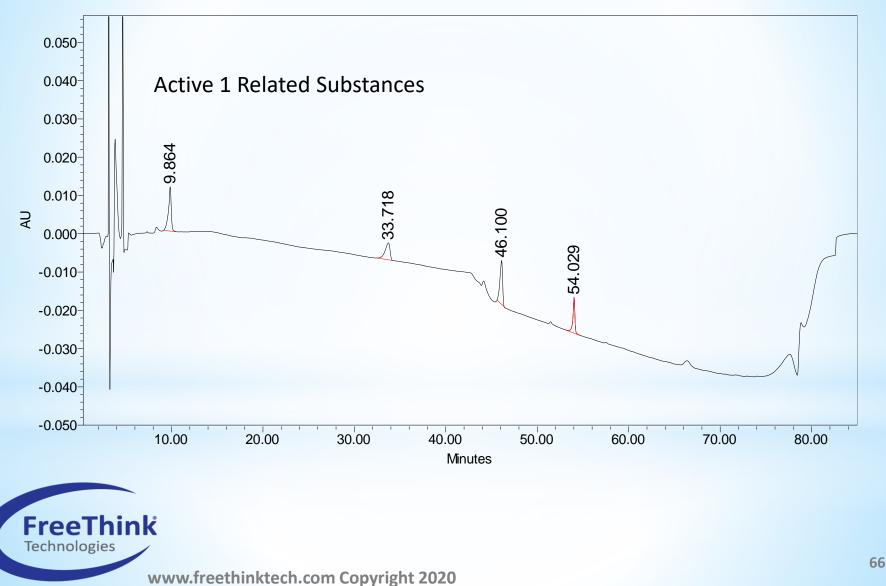


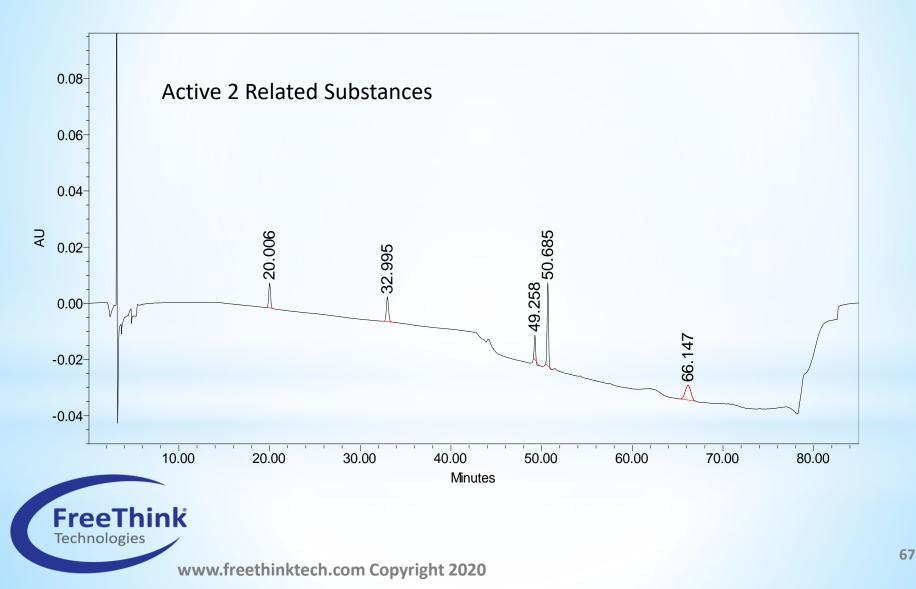
### **Final separation conditions:**

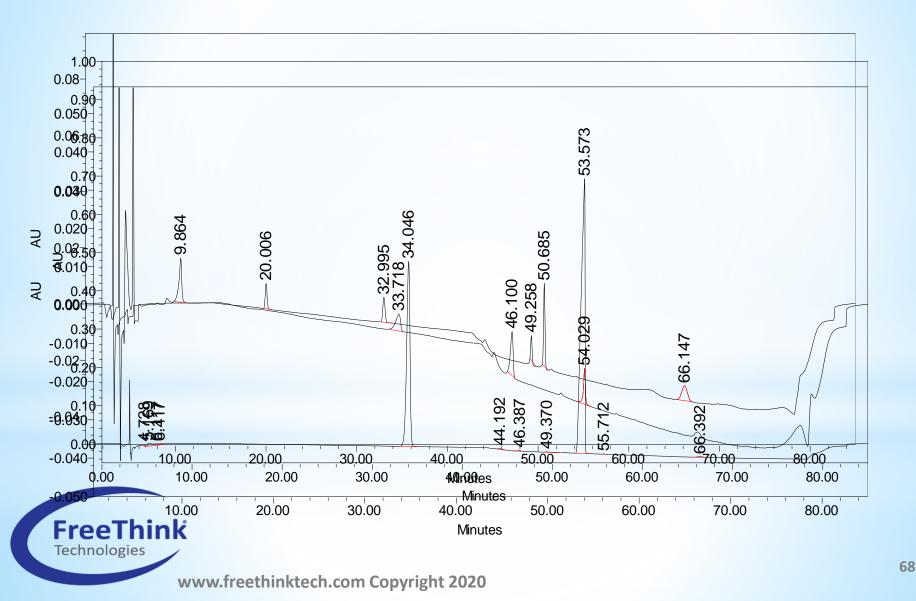
 Column selected was ABZ plus – see chromatograms on following slides



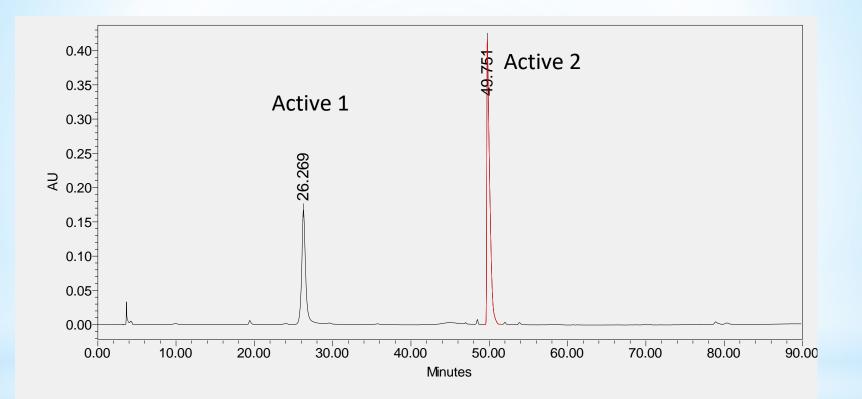






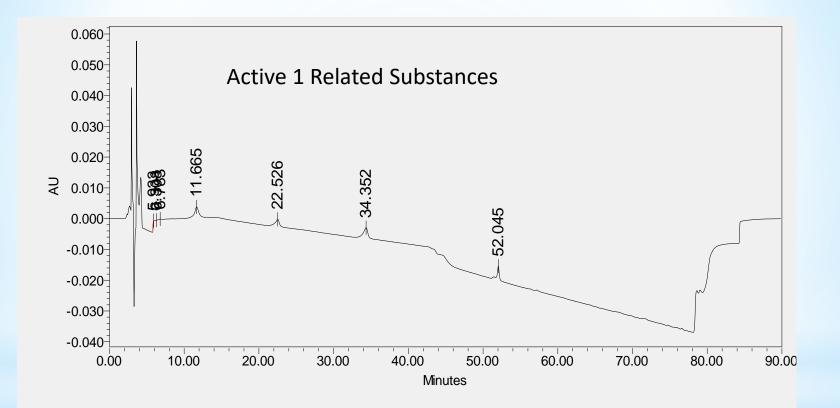


#### **ABZ Plus Column**



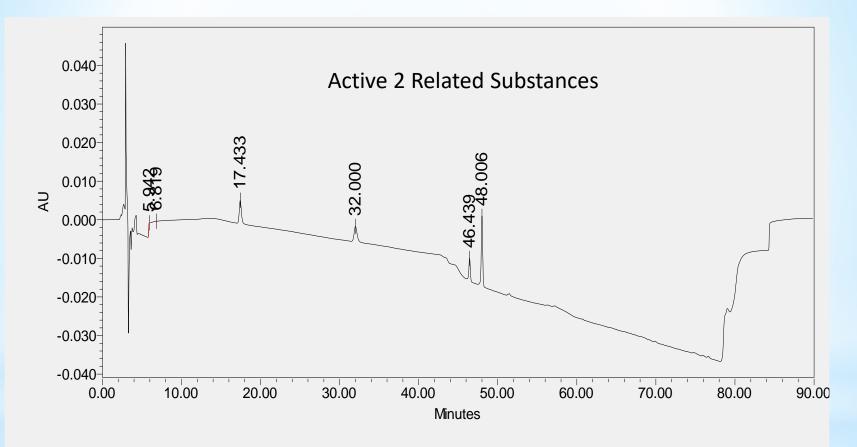


#### **ABZ Plus Column**



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#### **ABZ Plus Column**



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